

## Nickel-Containing Hydrogenases and CO Dehydrogenases

Yang Zhang

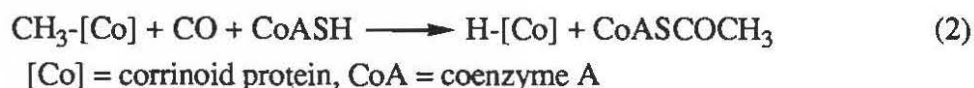
Literature Seminar

April 8, 1993

Nickel-containing enzymes have attracted attention as biological catalysts involved in several reactions that are of scientific importance. These enzymes are usually isolated from bacteria and algae. Four types of Ni-containing enzymes are now known [1]: ureases, methyl-coenzyme M reductases, hydrogenases and carbon monoxide dehydrogenases. The latter two types will be discussed in detail.

Hydrogenases catalyze the reversible oxidation of  $H_2$  to  $H^+$ . Several Ni-containing hydrogenases have been characterized, and they differ considerably in their active site composition [2]. They consist of two subunits of approximately 60 and 30 KDa and contain one or several [4Fe-4S] clusters plus one Ni atom. Hydrogenases are typically isolated in air as a mixture of inactive forms with distinct EPR spectra. The so-called Form A is only slowly reactivated after incubation under  $H_2$ . Form B is also inactive but is readily activated upon exposure to  $H_2$ . Enzyme reactivation under  $H_2$  leads to a new EPR-detectable species (Form C). These EPR signals are attributed to the Ni center by  $^{61}Ni$  substitution experiments [3]. EPR, ENDOR spectroscopy and XAS have been used to investigate the Ni active site environment of the hydrogenases [4]. It has been suggested that the Ni site features a distorted pseudo-octahedral structure. EXAFS data indicate that the Ni center is surrounded by 2 or 3 S ligands at an average Ni-S distance of 2.21 Å and 2 to 4 N or O ligands at 2.06 Å. A Ni-Fe distance  $>4$  Å is also indicated [5]. A tentative catalytic and activation scheme has been proposed showing the involvement of the redox centers in the hydrogenases [6]. Two Ni compounds with  $S_4N_2$  and  $S_2N_3$  coordination spheres have been synthesized as structure models of these hydrogenases [7].

The anaerobic CO dehydrogenases (CODH) catalyze the following reactions:



Reaction 2 is a sum of the final steps in CO fixation via acetyl-CoA synthesis. CODH is the central participant in this reaction [8]. The pathway responsible for reaction (2) involves several additional enzymes. The mechanism shown in Fig. 1 has been proposed by Wood et al. [9]. CODH from *C. thermoaceticum* has been determined to have an  $(\alpha\beta)_3$  hexameric protein subunit structure, where each  $\alpha\beta$  unit consists of 2 Ni, 11 Fe, 1 Zn and 31 S atoms [10]. The as-isolated CODH from *C. thermoaceticum* is EPR silent. Incubation under CO leads to an axial EPR spectrum with g values of 2.08 and 2.02. This EPR signal arises from a species containing Ni, Fe and the carbon from CO, as demonstrated by the hyperfine broadening of the signals when either  $^{61}Ni$ ,  $^{13}C$ , or  $^{57}Fe$  is present [11]. The complex responsible for this EPR signal has been referred to as a NiFeC complex. The CODHs from other bacteria exhibit similar but not identical EPR spectra. EPR, Mössbauer and ENDOR data [12] on the CO-reduced form of the enzyme indicate a Ni-Fe-C assembly of probable stoichiometry  $Ni_1Fe_{3-4}S_{2-4}C_1$ . EXAFS results on the CO-free, EPR silent form of the enzyme reveal a sulfur-rich nickel coordination environment, possibly planar  $NiS_4$  with Ni-S

distance of 2.16 Å. A combination of S/O/N ligands may also exist [13]. A N, O, S-ligated Ni complex and complexes with composition of  $[\text{Ni}(\text{NCH}_2\text{CH}_2\text{SR})\text{L}]^+$  (L = Cl, H, CO; R = i-Pr, t-Bu) have been used as models of CODHs [14].

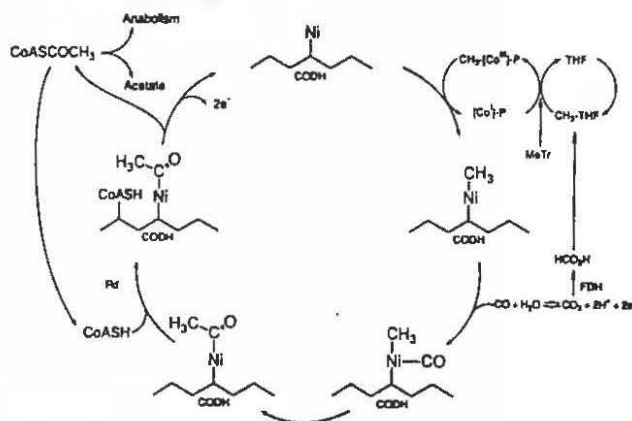


Figure 1. Proposed reaction pathway of Ni-CODH as an acetyl-coenzyme A synthase: CoA = coenzyme A, FDH = formate dehydrogenase, MeTr = methyltransferase, [Co]-P = corrinoid protein, Rd = disulfide reductase, THF = tetrahydrofolate.

Only preliminary results have been obtained in this area. The exact electronic and molecular structures of these enzymes remain unresolved and the catalytic roles of the active sites need to be established.

## References

1. For a general review of Ni-containing enzymes, see *The Bioinorganic Chemistry of Nickel*; Lancaster, J. R., Ed.; VCH Publishers, Inc.: New York, 1988.
2. Cammack, R.; Fernandez, V. M.; Schneider, K.; Chap. 8 in ref. 1.
3. Moura, J.; Moura, I.; Huynh, B. H.; DuVarney, R. C.; Xavier, A. V.; Peck, H. D.; LeGall, J., "Unambiguous Identification of The Nickel EPR Signal in  $^{61}\text{Ni}$ -Enriched *D. gigas* Hydrogenase," *Biochem. Biophys. Res. Commun.* **1982**, *108*, 1388.
4. (a) Trixeria, M.; Moura, I.; Xavier, A. V.; LeGall, J.; DerVartanian, D. V.; Peck, H. D.; Huynh, B. H., "Redox Intermediates of *D. gigas* [NiFe] Hydrogenase Generated Under Hydrogen," *J. Biol. Chem.* **1989**, *264*, 16435.  
 (b) Cammack, R.; Bagyinka, C.; Kovacs, K. L., "Spectroscopic Characterization of the Nickel and Iron-Sulfur Clusters of Hydrogenase from the Purple Photosynthetic Bacterium *T. roseopersicina*," *J. Biochem.* **1989**, *182*, 357.  
 (c) Scott, R. A.; Wallin, S. A.; Czechowski, M.; DerVartanian, D. V.; LeGall, J.; Peck, H. D.; Moura, I., "X-ray Absorption Spectroscopy of Nickel in the Hydrogenase from *D. gigas*," *J. Am. Chem. Soc.* **1984**, *106*, 6864.  
 (d) Whitehead, J. P.; Colpas, G. J.; Bagyinka, C.; Maroney, M. J., "X-ray Absorption Spectroscopic Study of the Reductive Activation of *T. roseopersicina* Hydrogenase," *J. Am. Chem. Soc.* **1991**, *113*, 6288.

- (e) Fan, C.; Teixeira, M.; Moura, J.; Moura, I.; Huynh, B. H.; Hoffman, B. M., "Detection and Characterization of Exchangeable Protons Bound to the Hydrogen-Activation Nickel Site of *D. gigas* Hydrogenase: A  $^1\text{H}$  and  $^2\text{H}$  Q-Band ENDOR Study," *J. Am. Chem. Soc.* **1991**, *113*, 20.
5. Maroney, M. J.; Colpas, G. J.; Bagyinka, C.; Baidya, N.; Mascharak, P. K., "EXAFS Investigation of the Ni Site in *T. roseopersicina* Hydrogenase: Evidence for a Novel Ni, Fe, S. Cluster," *J. Am. Chem. Soc.* **1991**, *113*, 3962.
6. Trixeria, M.; Moura, E.; Xavier, A. V.; LeGall, J.; DerVartanian, D. V.; Peck, H. D.; Huynh, B. H.; Moura, J. G., "EPR Studies on the Mechanism of Activation and the Catalytic Cycle of the Nickel-containing Hydrogenase from *D. gigas*," *J. Biol. Chem.* **1985**, *260*, 8942.
7. (a) Baidya, N.; Olmstead, M. M.; Whitehead, J. P.; Bagyinka, C.; Maroney, M. J.; Mascharak, P. K., "X-ray Absorption Spectra of Nickel Complexes with  $\text{N}_3\text{S}_2$  Chromophores and Spectroscopic Studies on  $\text{H}^-$  and CO Binding at These Nickel Centers: Relevance to the Reactivity of the Nickel Site(s) in [FeNi] Hydrogenase," *Inorg. Chem.* **1992**, *31*, 3612.  
(b) Kruger, H. J.; Holm, R. H., "Stabilization of Trivalent Nickel in Tetragonal  $\text{NiS}_4\text{N}_2$  and  $\text{NiN}_6$  Environments: Synthesis, Structure, Redox Potentials, and Observations Related to [NiFe]-Hydrogenases," *J. Am. Chem. Soc.* **1990**, *112*, 2963.
8. Ragsdale, S. W.; Wood, H. G., "Acetate Biosynthesis by Acetogenic Bacteria," *J. Biol. Chem.* **1985**, *260*, 3970.
9. Pezacka, E.; Wood, H. G., "The Autotrophic Pathway of Acetogenic Bacteria," *J. Biol. Chem.* **1986**, *261*, 1609.
10. Ragsdale, S. W.; Clark, J. E.; Ljungdahl, L. G.; Lundie, L. L.; Drake, H. L., "Properties of Purified Carbon Monoxide Dehydrogenase from *C. thermoacetikum*, a Nickel, Iron-Sulfur Protein," *J. Biol. Chem.* **1983**, *258*, 2364.
11. Ragsdale, S. W.; Wood, H. G.; Antholine, W. E., "Evidence that an Iron-Nickel-Carbon Complex is Formed by Reaction of CO with the CO Dehydrogenase from *C. thermoacetikum*," *Proc. Natl. Acad. Sci. USA*, **1985**, *82*, 6811.
12. (a) Lindahl, P. A.; Munck, E.; Ragsdale, S. W., "CO Dehydrogenase from *C. thermoacetikum*," *J. Biol. Chem.* **1990**, *265*, 3873.  
(b) Lindahl, P. A.; Ragsdale, S. W.; Munck, E., "Mössbauer Study of CO Dehydrogenase from *C. thermoacetikum*," *J. Biol. Chem.* **1990**, *265*, 3880.  
(c) Fan, C.; Gorst, C. M.; Ragsdale, S. W.; Hoffman, B. M., "Characterization of Ni-Fe-C Complex Formed by Reaction of Carbon Monoxide with the Carbon Monoxide Dehydrogenase from *C. thermoacetikum* by Q-Band ENDOR," *Biochem.* **1991**, *30*, 431.
13. (a) Tan, G. O.; Ensign, S. A.; Scott, M. J.; Holm, R. H.; Stephens, P. J.; Hodgson, K. O., "On the Structure of the Ni/Fe/S Center of the CO Dehydrogenase from *R. rubrum*: An X-ray Absorption Spectroscopy Study," *Proc. Natl. Acad. Sci. USA* **1992**, *89*, 4427.

- (b) Crammer, S. P.; Eidesness, M. K.; Pan, W.-H.; Morton, T. A.; Ragsdale, S. W.; DerVartanian, D. V.; Ljungdahl, L. G.; Scott, R. S., "X-ray Absorption Spectroscopy Evidence for a Unique Nickel Site in *C. thermoaceticum* CO Dehydrogenase," *Inorg. Chem.* **1987**, *26*, 2477.
  - (c) Bastian, N. R.; Diekert, G.; Niederhoffer, E. C.; Teo, B.-K.; Walsh, C. T.; Orme-Johnson, W. H., "Nickel and Iron EXAFS of CO Dehydrogenase from *C. thermoaceticum* Strain," *J. Am. Chem. Soc.* **1988**, *110*, 5581.
- 14.
- (a) Stavropoulos, P.; Muetterties, M. C.; Carrie, M.; Holm, R. H., "Structural and Reaction Chemistry of Nickel Complexes in Relation to CO Dehydrogenase: A Reaction System Simulating Acetyl-Coenzyme A Synthase Activity," *J. Am. Chem. Soc.* **1991**, *113*, 8485.
  - (b) Lu, Z.; White, C.; Rheingold, A. L.; Crabtree, R. H., "Functional Modeling of CO Dehydrogenase: Catalytic Reduction of Methylviologen by CO/H<sub>2</sub>O with N, O, S-Ligated Nickel Catalyst," *Angew Chem. Int. Ed. Engl.* **1993**, *32*, 92.